

Identification of *act2*, an essential gene in the fission yeast *Schizosaccharomyces pombe* that encodes a protein related to actin

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ABSTRACT Actins are a family of highly conserved proteins that are ubiquitously found among eukaryotic organisms. All actins that have previously been identified, including those of animals, plants, fungi, and protozoa, are 374–376 amino acids long and exhibit at least 70% amino acid sequence identity when compared with one another. We have cloned a gene from the fission yeast *Schizosaccharomyces pombe* that encodes a distantly related member of the actin protein family, herein referred to as *act2*. In contrast to all other actins, the derived amino acid sequence reveals that *act2* is 427 residues long and exhibits only 35–40% identity to actins, including *act1* from *Sch. pombe*. Comparison to the known x-ray crystallographic structure of rabbit skeletal muscle actin indicates that the ATP and divalent metal ion binding sites are largely conserved in *act2*, while regions involved in actin–actin and actin–myosin interactions are relatively divergent. Disruption of the *act2* gene demonstrated that this gene encodes a function essential for germination of haploid spores. These findings indicate that while *act2* and *act1* are related proteins, they appear to have distinct functions. In addition, they demonstrate that the actin protein family is more diverse than was previously thought.

The shape and motility of eukaryotic cells are largely governed by three filamentous systems—actin filaments, intermediate filaments, and microtubules. Each of these systems possesses a major core protein—namely, actin, intermediate filament protein, and tubulin, respectively. In a given organism or cell type, considerable isoform diversity of both core proteins and associated proteins is often present. Actin filaments consist of a double-helical array of actin monomers polymerized in a polar manner. A number of proteins are associated with actin that function in nucleating, capping, stabilizing, severing, bundling, and mechanical movement of actin filaments. Through the interactions with these proteins, actin filaments participate in several essential cellular functions, including cell motility, cytokinesis, maintenance of cell structure, and organelle movement (reviewed in ref. 1). Many organisms have multiple isoforms of actin that often exhibit developmentally regulated and cell-type-specific expression (reviewed in refs. 2 and 3). Actin isoforms can also exhibit differential localization within a single cell (4). Actins that have been characterized from a broad range of phyla including vertebrates, invertebrates, fungi, plants, and protozoa have between 70% and 90% amino acid sequence identity and are 374–376 amino acids in length. Budding yeast (*Saccharomyces cerevisiae*) and fission yeast (*Schizosaccharomyces pombe*) each possess a single gene (*act1*) encoding an actin with an amino acid sequence that is ≈90% identical to the sequence of vertebrate actins (5, 6). The *act1* gene of budding yeast is known to be essential for viability (7). Several proteins that interact with actin have been identified in budding yeast, including profilin, tropomyosin, and myosin

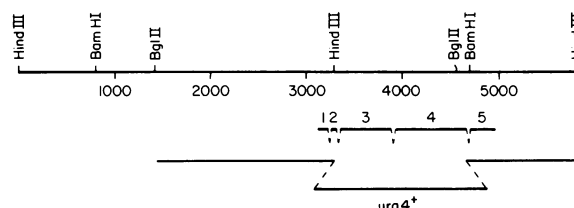


FIG. 1. Restriction map of the 5.8-kilobase (kb) *HindIII*–*HindIII* fragment carrying *act2* is shown with the 5′-end to the left (*Top*). The localization of the five exons (*Middle*) and the region deleted for gene disruption (*Bottom*) are illustrated. To make the disruption construct, an internal 2.5-kb *HindIII*–*Bam*HI fragment and the 3′-end 1.1-kb *Bam*HI–*HindIII* fragment were each inserted into a pUC19 vector. The vector containing the internal fragment was cut with *Hind*III, a *Bam*HI linker was added destroying the *Hind*III site, and the construct was digested with *Bam*HI and *Bgl* II. The resulting 1.9-kb fragment was ligated into the *Bam*HI site of the vector containing the 3′-end fragment so as to leave an intact internal *Bam*HI site into which the *ura4*⁺ gene was inserted. These manipulations resulted in the replacement of a 1.4-kb *Hind*III–*Bam*HI fragment containing 90% of the *act2* gene coding sequence with a 1.8-kb fragment containing the *ura4*⁺ gene (13). A *h*⁺*N*/*h*⁺*N* *ura4*–*D18/ura4*–*D18 ade6*–*210/ade6*–*216 leu1*–*32/leu1*–*32* diploid strain (SP826) was transformed to *Ura*⁺ with a 4.8-kb *Eco*RI–*Hind*III fragment excised from this plasmid. Southern hybridization to DNA from the transformants indicated that the appropriate homologous recombination had occurred in 10 of 16 cases. A *h*⁺*N*/*h*⁹⁰ strain capable of high-frequency sporulation was selected from one of the original *act2*::*ura4*⁺ recombinants by iodine staining of replica plates and was used for tetrad analysis.

(reviewed in ref. 8). Actin in yeast is localized in both patches and cables, the former being associated with regions of cell growth including the bud neck of budding yeast and the cytokinetic septum of fission yeast (8–10). In this study we have characterized a yeast gene (*act2*) encoding an actin-like protein that, at present, is the most distantly related member of the known actin gene family.†

MATERIALS AND METHODS

Cloning of *act2*. A *Sch. pombe* cDNA library in Lambda ZAP (Stratagene) was kindly provided by Dallan Young (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY). Oligodeoxynucleotides YTM.15 (5′-AAAGGCAACCATG-GACAAAT-3′) and YTM.13 (5′-ACAAAAGTTGATCA-CAAGTT-3′) were synthesized with the intention of cloning a yeast tropomyosin cDNA by amplification with the polymerase chain reaction (11). DNA amplification mixtures (50 μl) consisted of 1 μg of cDNA library, 50 pmol of each primer, buffer, dNTPs, and *Taq* polymerase as recommended by Perkin–Elmer/Cetus and were incubated for 35 cycles of 1.5 min at 94°C, 3 min at 45°C, and 2 min at 72°C. A 660-nucleotide fragment was obtained that, when cloned into pUC19 and sequenced with Sequenase (United States Bio-

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†The sequence reported in this paper has been deposited in the GenBank data base (accession no. M81068).

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aattctcttttcgcttaccttttcataaaataatagcgca ATG GCT TCG TTT AAT GTT CCC ATT ATA ATG 30
Met Ala Ser Phe Asn Val Pro Ile Ile Met 10

GAC AA gtaagtattgatgtagcgaagttaaattgttcgtgaatatcagttactaactttatttgttagC GGT ACC 105
Asp Asn Gly Thr

GGC TAT AGC AAG CTT GG gtatgtgtaaatagtcgttttaggatgttattatctctcttttgagcattttaaac 178
Gly Tyr Ser Lys Leu Gly 20

agtaatttaatttcag C TAT GCT GGG AAC GAT GCT CCT TCT TAT GTA TTC CCC ACT GTT ATT 240
Tyr Ala Gly Asn Asp Ala Pro Ser Tyr Val Phe Pro Thr Val Ile 35

GCG ACG CGT TCT GCG GGT GCT TCT TCA GGG CCG GCT GTT TCA TCA AAG CCT TCT TAC ATG 300
Ala Thr Arg Ser Ala Gly Ala Ser Ser Gly Pro Ala Val Ser Ser Lys Pro Ser Tyr Met 55

GCC TCG AAA GGC AGT GGG CAT CTC TCA AGC AAA AGG GCA ACT GAG GAC TTG GAC TTT TTT 360
Ala Ser Lys Gly Ser Gly His Leu Ser Ser Lys Arg Ala Thr Glu Asp Leu Asp Phe Phe 75

ATT GGA AAC GAT GCA TTG AAA AAG GCG TCT GCT GGA TAT TCT TTG GAC TAT CCT ATC CGT 420
Ile Gly Asn Asp Ala Leu Lys Lys Ala Ser Ala Ser Leu Asp Tyr Pro Ile Arg 95

CAT GGA CAA ATA GAA AAT TGG GAC CAT ATG GAG CGT TTT TGG CAA CAG TCT CTC TTT AAA 480
His Gly Gln Ile Glu Asn Trp Asp His Met Glu Arg Phe Trp Gln Gln Ser Leu Phe Lys 115

TAT TTA CGT TGT GAA CCA GAG GAC CAT TAT TTT CTT TTG ACC GAA CCT CCT TTG AAT CCA 540
Tyr Leu Arg Cys Glu Pro Glu Asp His Tyr Phe Leu Leu Thr Glu Pro Pro Leu Asn Pro 135

CCT GAA AAC CGT GAA AAT ACT GCA GAG ATT ATG TTT GAG TCG TTC AAC TGT GCA GGA CTC 600
Pro Glu Asn Arg Glu Asn Thr Ala Glu Ile Met Phe Glu Ser Phe Asn Cys Ala Gly Leu 155

TAC ATT GCT GTT CAA GCG GTT TTG GCT CTT GCC GCC TCT TGG ACA TCT TCA AAA GTT ACT 660
Tyr Ile Ala Val Gln Ala Val Leu Ala Ala Ser Trp Thr Ser Ser Lys Val Thr 175

GAT CGT TCG TTG ACC GGT ACT GTT GTT GAC TCT GGT GAT GGC GTT ACT CAT ATT ATT CCT 720
Asp Arg Ser Leu Thr Gly Thr Val Val Asp Ser Gly Asp Gly Val Thr His Ile Ile Pro 195

GTT gtatgtataaaaaatctaatacagcgttagataaacagttttgaacattaaatgcatatatatgcccgaattt 798
Val Val 196

tactaactgataaaccttagGCC GAA GGA TAT GTT ATC GGC TCT TCC ATT AAG ACA ATG CCA CTT 863
Ala Glu Gly Tyr Val Ile Gly Ser Ser Ile Lys Thr Met Pro Leu 211

GCT GGT CGT GAT GTT ACG TAC TTT GTT CAA TCC TTA TTA CGT GAC CGC AAT GAA CCT GAT 923
Ala Gly Arg Asp Val Thr Tyr Phe Val Gln Ser Leu Leu Arg Asp Arg Asn Glu Pro Asp 231

TCA AGT CTC AAA ACT GCT GAA AGA ATT AAA GAG GAA TGT TGC TAC GTT TGT CCG GAC ATT 983
Ser Ser Leu Lys Thr Ala Glu Arg Ile Lys Glu Glu Cys Cys Tyr Val Cys Pro Asp Ile 251

GTT AAG GAG TTT AGT AGA TTT GAT CGT GAG CCC GAC CGT TAT TTA AAA TAT GCA TCT GAA 1043
Val Lys Glu Phe Ser Arg Phe Asp Arg Glu Pro Asp Arg Tyr Leu Lys Tyr Ala Ser Glu 271

TCA ATT ACT GGA CAC TCT ACT ACT ATT GAT GTA GGA TTT GAA CGC TTC CTC GCA CCT GAA 1103
Ser Ile Thr Gly His Ser Thr Thr Ile Asp Val Gly Phe Glu Arg Phe Leu Ala Pro Glu 291

ATA TTT TTT AAT CCT GAG ATT GCT TCT TCC GAT TTT TTG ACC CCT CTT CCG GAA CTT GTG 1163
Ile Phe Phe Asn Pro Glu Ile Ala Ser Ser Asp Phe Leu Thr Pro Leu Pro Glu Leu Val 311

GAC AAC GTC GTA CAG AGC TCA CCC ATT GAC GTT CGT AAA GGC CTT TAT AAA AAC ATT GTC 1223
Asp Asn Val Val Gln Ser Ser Pro Ile Asp Val Arg Lys Gly Leu Tyr Lys Asn Ile Val 331

CTT TCT GGT GGT AGT ACT TTG TTT AAG AAC TTT GGC AAC CGC TTA CAA AGA GAT CTC AAG 1283
Leu Ser Gly Gly Ser Thr Leu Phe Lys Asn Phe Gly Asn Arg Leu Gln Arg Asp Leu Lys 351

CGG ATT GTT GAC GAG CGT ATT CAC CGC AGT GAA ATG CTC TCC GGT GCT AAA AGT GGT GGT 1343
Arg Ile Val Asp Glu Arg Ile His Arg Ser Glu Met Leu Ser Gly Ala Lys Ser Gly Gly 371

GTA GAC GTT AAT GTT ATC TCC CAT AAA CGT CAA CGA AAT GCG GTT TGG TTT GGC GGC AGT 1403
Val Asp Val Asn Val Ile Ser His Lys Arg Gln Arg Asn Ala Val Trp Phe Gly Gly Ser 391

TTA CTA GCA CAA ACG gtatgtcattatcttattgtttgtatatatttttagtttaccgtagagatttactgacat 1477
Leu Leu Ala Gln Thr 396

gtttag CCT GAA TTC GGA TCC TAT TGC CAT ACC AAA GCA GAT TAC GAA GAA TAT GGT GCT 1537
Pro Glu Phe Gly Ser Tyr Cys His Thr Lys Ala Asp Tyr Glu Glu Tyr Gly Ala 414

TCC ATT GCT CGT AGG TAC CAA ATT TTT GGA AAT TCT CTT taatgaaatatcatgagttttgcgt 1603
Ser Ile Ala Arg Arg Tyr Gln Ile Phe Gly Asn Ser Leu 427

ctataggatttactgattaaaatgaaaagtttaatggagcaggcattagccttagaagaaatagacttgaggtttgcta 1682

tgtttccaggctttttcattgagcaatatcaaataagatgtatataattaacttgctc 1741

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FIG. 2. Nucleotide sequence and encoded amino acid sequence of the *act2* gene. The gene possesses five exons that encode a total of 427 amino acids. The four introns are written in lowercase type, and the branch point and splicing signals are underlined.

chemical), was found to encode an amino acid sequence similar to that of actin. When the sequence of the full-length *act2* cDNA (obtained through plaque hybridization from the above library) was later determined, both oligonucleotides were found to have primed out of frame so that no similarity between actin and tropomyosin should be inferred. The *act2* gene was selected through colony hybridization from a *Sch. pombe* genomic library in the pWH5 vector (12).

RESULTS AND DISCUSSION

We first identified a cDNA encoding part of an actin-like protein sequence through selective amplification of a *Sch. pombe* cDNA library with the polymerase chain reaction (11). The partial cDNA was cloned into the pUC19 vector and used to detect a full-length cDNA from the same library. This cDNA was then used to screen a *Sch. pombe* genomic library in the plasmid vector pWH5 (12) (Fig. 1). The comparison of

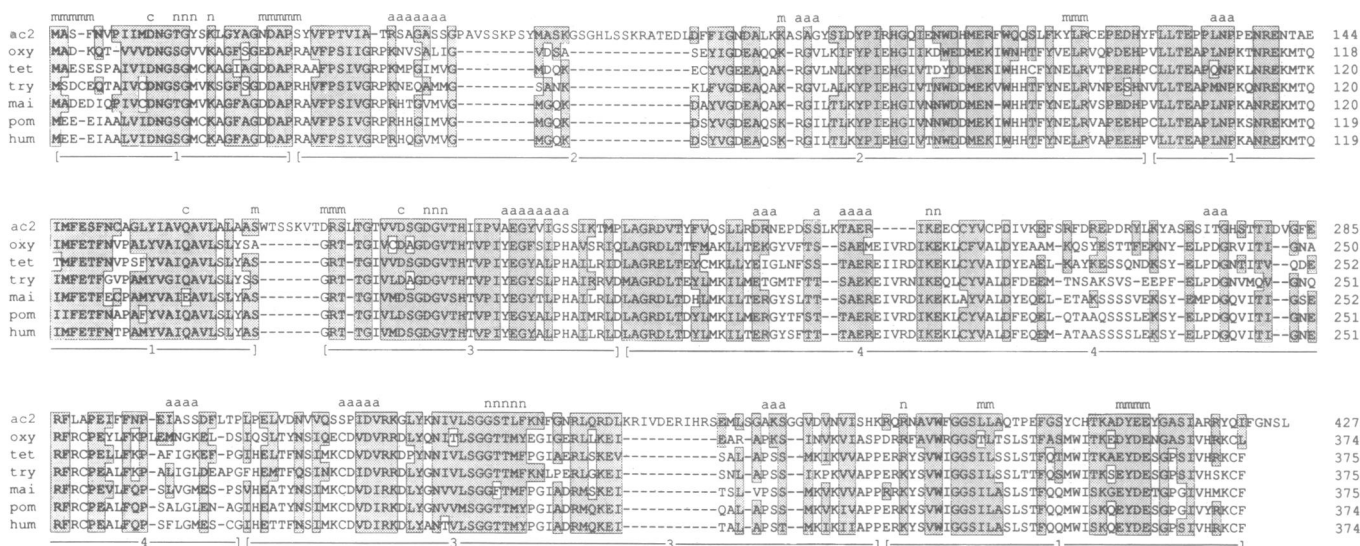


FIG. 3. Comparison of the amino acid sequences of act2 (ac2) and actins from diverse species: The original alignments were made with FASTDB using a McLachlan structure genetic matrix and a window size of 60 (15). Minor modifications were made manually. Amino acids are depicted by the conventional one-letter code. Residues that are similar or identical to the act2 sequence are shaded. Similarity is defined as (E/D), (N/Q), (K/R), (A/G), (S/T), (Y/F), and (L/I/M/V). The residues in actin that interact with the nucleotide ATP (n), Ca^{2+} (c), and myosin (m) and that are involved in actin-actin interactions (a) as determined by x-ray crystallography and other methods (16, 17) are indicated above the act2 sequence. The regions corresponding to the four subdomains of actin are indicated below the actin sequences. oxy, *Oxytricha nova* macronuclear actin (18); tet, *Tetrahymena thermophila* actin (19); try, *Trypanosoma brucei* actin (20); mai, maize actin (21); pom, *Sch. pombe* actin (6); hum, human cytoplasmic γ -actin (22). Gaps required for alignment are indicated by dashes.

cDNA and genomic sequences revealed the presence of four introns in the *act2* gene (Fig. 2). Introns 1, 3, and 4 have sequences that match the consensus for 5' and 3' splice sites and the branch point in *Sch. pombe* (14). Intron 2 is unusual in having a T in the first position of the CTA/GAC/T branch-point consensus sequence. The *act2* introns are located in the codons for amino acids Asn-12 and Gly-20, and after codons encoding for Val-196 and Thr-396. According to our alignment (Fig. 3), the corresponding positions in actins are at amino acids 12, 20, 164, and 349. Although a total of >30 intron positions have been described in actin genes from a variety of organisms, none is equivalent to those found in the *act2* gene (23). Dibb and Newman (24) have proposed that most introns were not present in the ancestral actin gene but have been gained during the evolution of the actin gene family through insertion into a consensus sequence, C/AAGR. While the *act2* introns do not appear to have inserted within this sequence, the growing number of intron positions in actin genes supports the hypothesis that insertions are likely to have occurred. From analysis of multiple cDNAs and Northern blotting (data not shown), two alternative polyadenylation sites were identified in the *act2* gene, 26 and 165 base pairs downstream of the UAA stop codon. The majority of *act2* mRNAs use the downstream site.

In order to determine whether or not *act2* is essential for viability, a null allele of the gene was obtained by the one-step gene disruption method (25) (Fig. 1). Following gene disruption in the diploid strain SP826, tetrad analysis of 30 asci revealed 24 cases of 2:2 segregation where two spores developed on Ura⁻ colonies and two spores failed to germinate, 5 cases with one colony, and 1 with three colonies. No Ura⁺ colonies were recovered from any of the tetrads analyzed. These results indicate that the presence of an intact *act2* gene is essential for germination to occur in *Sch. pombe*. In addition, they indicate that act2 is likely to have a function distinct from that of act1. They do not, however, eliminate the remote possibility that overexpression of act1 may compensate for loss of act2 expression.

The *act2* cDNA has a 427-amino acid open reading frame that encodes a protein with a calculated M_r of 47,376 and pI

of 5.7, compared with a typical actin with M_r of 42,000 and pI of 5.2. We have estimated the abundance of act2 through codon usage analysis. In *Sch. pombe*, highly expressed proteins tend to be encoded by different and fewer codon types than low-abundance proteins. These differences are measured in terms of a codon bias index (CBI) (26). We analyzed the codon bias of both *act1* and *act2* genes from *Sch. pombe* and found that the *act1* gene (CBI = 0.70) had a value similar to those of genes encoding highly expressed enzymes such as triose-phosphate isomerase, while *act2* (CBI = 0.18) was similar to genes encoding regulatory proteins such as cdc25 and wee1. These results indicate that act2 is unlikely to represent a large portion of the actin-based structures detected in yeast by phalloidin or by anti-actin antibodies (10).

When the act2 amino acid sequence was compared with the National Biomedical Research Foundation Protein Identification Resource (version 24) and GenBank (version 64) data bases, the large group of actin sequences possessed alignment scores significantly above any other in the data base (Fig. 3). Amino acid identity to actins ranged from 35% to 40% (based on the length of actin) whereas similarity as defined in Fig. 3 was 50–55%. The act2 protein was no more similar to *Sch. pombe* actin (act1) than to actins from a variety of other organisms, supporting the possibility that it diverged from actin early in eukaryotic evolution and will be found in other species. The protozoan actins are among the most divergent, possessing only 70–75% amino acid sequence identity to actins from animals, plants, and fungi. However, they have the same level of identity with act2 (35–40%) as the other actins. This observation is also in agreement with an ancient divergence of the *act2* and actin genes.

Although act2 and actin have clearly evolved from a common ancestral gene, it remains to be determined whether these proteins share any functional properties. The elucidation of the x-ray crystallographic structure of rabbit skeletal muscle actin (16) has allowed us to make some theoretical predictions regarding possible functional similarities and differences between these proteins. Actin possesses two

domains each containing two subdomains. Subdomains 1 and 3 each contain a five-stranded β -sheet that includes a $\beta\alpha\beta$ unit. These structures are largely responsible for the ability of actin to bind ATP and divalent metal cation. Alignment of the amino acid sequence of act2 with that of actin indicates that residues which interact with ATP and metal ion are highly conserved with the possible exception of residue 16, which has a substitution of tyrosine for methionine. The essential structural features of subdomains 1 and 3 also appear to be conserved, indicating that act2 is likely to possess ATPase activity (Fig. 3). The residues thought to be involved in actin-actin contacts (17) are generally not highly conserved in act2. Eight of nine predicted actin-actin interaction sites exhibit at least one nonconservative substitution or insertion in act2 and, in particular, a large insertion near the amino terminus disrupts a region thought to be important in filament formation (Fig. 3). Other actins from diverse species, including that of *Tetrahymena*, which copolymerizes with vertebrate muscle actin (27), have far greater conservation of residues necessary for actin-actin interaction. These observations do not, however, rule out the possibility that act2 is incorporated into filaments. Among the sites in actin that are thought to interact with myosin, those near the carboxyl terminus are conserved in act2, whereas the amino-terminal sites are divergent. It will be interesting to determine whether act2 interacts with a myosin isoform.

Several proteins that interact with actin, including the capping/severing proteins gelsolin, villin, fragmin, and severin, share short regions of sequence similarity with actin (reviewed in ref. 28). While act2 is considerably more similar to actin than these proteins, its apparent low abundance makes it a candidate for an actin capping/severing factor. It is also possible that act2 has a function independent of actin. Despite lacking obvious sequence similarity, actin and the ATPase domain of 70-kDa heat shock cognate protein have nearly identical structures (29). It is thought that these proteins, along with hexokinase, may define a class of proteins that use the energy generated from cleavage of ATP to drive conformational change.

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